

Pinking in White Wines – A Review

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In the late 1960s, a phenomenon was discovered in white wines. It was noted that certain white wines turned pink in the bottle. This phenomenon was dubbed as pinking. Research was done on the pinking to establish its cause and effect. Analysis of SO₂, pH and polyvinyl polypyrrolidone (PVPP) showed that a minimum of 45 mg/L of SO₂ were needed for the wine not to be susceptible to pinking. Tests on the decrease in pH showed that there was no increase in pink colour with a decrease in pH, which meant that monomeric anthocyanins were not the cause of pinking. Recent research claims that malvidin-3-O-glucoside is the most abundant monomeric anthocyanin found in pinked wines and could be the cause of pinking. This led to the theory that phenols contribute to pinking susceptibility, and this was accepted as fact in recent years. The establishment of a pinking assay in 1977 made the testing for pinking easier and cheaper for winemakers. The sales of PVPP increased as winemakers worked preventatively with their wine to decrease susceptibility to pinking. This review attempts to describe the history of pinking, the establishment of the assay, as well as to describe factors that could lead to pinking susceptibility in white wines.

INTRODUCTION

The first incidence of pinking in white wines was reported by Singleton and Esau in 1969. This led to a series of research articles on pinking from 1977 to 1983 by an Australian researcher, Dr Bob Simpson (1977a, 1977b, 1980a, 1980b; Simpson *et al.* 1982, 1983). This was followed by a research article on the use of polyvinyl polypyrrolidone (PVPP) by Lamuela-Raventós *et al.* (2001) and two articles on the presence of anthocyanins by Andrea-Silva *et al.* (2014) and Cosme *et al.* (2019). This literature review reports on the findings of Simpson and other researchers that investigated pinking susceptibility in white wines

Simpson (1977a) defines pinking as “the troublesome discolouration” that develops during the storage of white wines. He later adds that it develops over several days, but most likely after vinification or when the wine is no longer protected by a CO₂ blanket. This led to the discovery that pinking occurs after contact with air (Simpson, 1980a). In 1982, Simpson stated that “white wines develop a pink colouration on exposure to air”. Andrea-Silva *et al.* (2014) define pinking as “the appearance of a salmon-red blush in white bottled wines produced exclusively from white varieties”. Therefore, a comprehensive definition of pinking could be established, as follows: Pinking, or oxidative

pinking, is the slight discolouration of white wines from a pinkish to a salmon-red blush colour, affected by certain oenological processes before and after fermentation until storage during which the wine could come into contact with air.

Different cultivars have been reported to show some degrees of pinking susceptibility. In America, the white cultivars reported are Thompson Seedless, Semillon, Sauvignon blanc, Chardonnay and Chenin blanc (Tobe, 1983; Jones, 1989). In Australia, the cultivars reported to be prone to pinking are Muscat Gordo Blanco, Sultana, Palomino, Riesling, Doradillo and Crouchen (Simpson, 1977a). In Spain, the cultivars Sauvignon blanc, White Riesling, Chardonnay, Albariño, Macabeo, Xarel·lo, Parellada, Garnatxa blanca (Grenache) and Verdejo were reported to have the potential to pink (Lamuela-Raventós *et al.*, 2001). In Portugal, the cultivar reported was Síría (Andrea-Silva *et al.*, 2014), and in the Czech Republic it was Pinot blanc, Pálava, Pinot Gris, Sauvignon blanc, Grüner Veltliner and Chardonnay (unpublished data). This is an extensive list of white cultivars. According to Simpson (1977a, 1977b), Tobe (1983), Jones (1989), Lamuela-Raventós *et al.* (2001) and Andrea-Silva *et al.* (2014), the

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predominant cultivar that shows susceptibility to pinking is Sauvignon blanc. Winemakers must take note of this when producing Sauvignon blanc. Although these cultivars showed a tendency to pink, regional variations and yearly differences also influence the potential of these wines to pink (Simpson, 1977a; Andrea-Silva *et al.*, 2014).

Wines made by winemaking practices such as cooling of the must, cold fermentation and the use of inert gasses (Ar, N₂ and CO₂) show higher susceptibility to pinking (Singleton & Esau, 1969; Simpson *et al.*, 1982). This led to the assumption that air contact or O₂ increases pinking susceptibility (Simpson, 1980b). Other factors, such as storage temperatures, the presence of light, free SO₂ content and the pH of the wine also play a role in pinking susceptibility (Simpson, 1977; Simpson *et al.*, 1982). With the influence of light came the suggestion that wine must not be bottled in clear glass bottles, but rather in green or dark green bottles (Lamuela-Raventós *et al.*, 2001). Anecdotal evidence also suggests that pinking does not affect the aroma or taste of the white wines (Simpson, 1980b; Lamuela-Raventós *et al.*, 2001), but this has never been proven scientifically.

Simpson (1980b) states that there is “good evidence” that the compounds causing pinking have their origin in phenolics. This led to a worldwide belief that phenols cause pinking in white wines (Jacobson, 2006; Jackson, 2014, 2016), and that polyvinylpolypyrrolidone (PVPP) should be used for their removal (Lamuela-Raventós *et al.*, 2001).

MEASUREMENT OF PINK SUSCEPTIBILITY

Development of an assay

Simpson (1977a, 1980b) did extensive studies on the pinking susceptibility of white wines and the analysis thereof. Spectrophotometric studies on normal white wines and wines

with a visible pinking showed a distinctive bump over the 500 nm absorbency range (Figs 1 & 2). Therefore, because the greatest differences occurred at an optical density of 500 nm, this wavelength was chosen as a suitable wavelength for testing for pinking susceptibility.

The absorbance of a normal white wine therefore will have a smooth curve at 500 nm, but a white wine with a visible pink colouration will show an absorbency at 500 nm. Thus, when white wine is tested for pinking susceptibility, two samples of the wine are taken. One will be the control and the other the treatment. The control sample is determined first at 500 nm, followed by the pink induced sample. The difference between the two samples will show the pinking susceptibility of the wine. Simpson found that light-colour wines will show a pinking susceptibility of 5 (0.005 AU x 10³), and darker coloured wines will have a pinking susceptibility above 10 (0.01 AU x 10³). With darker coloured white wines, Simpson meant wines that border on a more yellowish colour.

Wines that show a tendency to browning rather than pinking will show a greater absorbency at 420 nm. At the wavelength of 420 nm, there will be no interference from the pink colouration. Both pinking and browning therefore can be measured in white wines.

Simpson (1977a) prepared a 0.3% (w/v) solution (1 mL in 100 mL distilled water) of 30% (w/v) hydrogen peroxide (H₂O₂). He used increments of 0.05 mL, starting from 0.05 mL in a 10 mL wine sample, and ending with 0.40 mL from this 0.3% (w/v) H₂O₂ solution to end up with concentrations ranging from 15 mg/L to 120 mg/L H₂O₂. For each wine tested, there were two samples, one of which was the control and the other one that received the H₂O₂ addition. These two samples of each wine were then kept in the dark for 24 hours before being analysed spectrophotometrically.

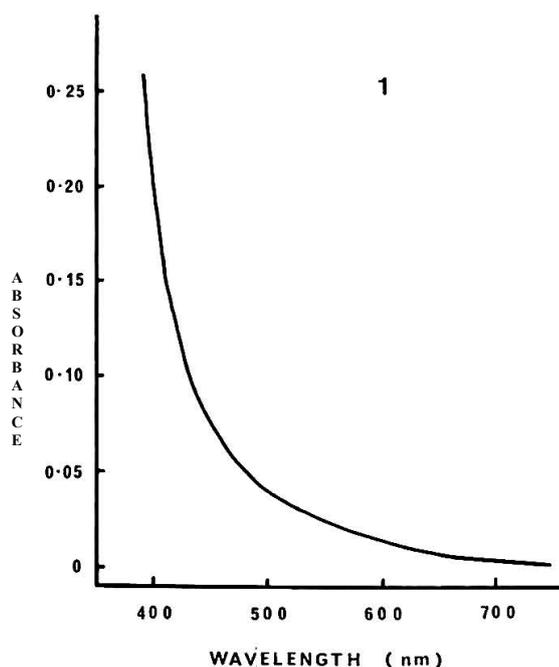


FIGURE 1

Spectrum of a wine showing no pinking (from Simpson, 1977).

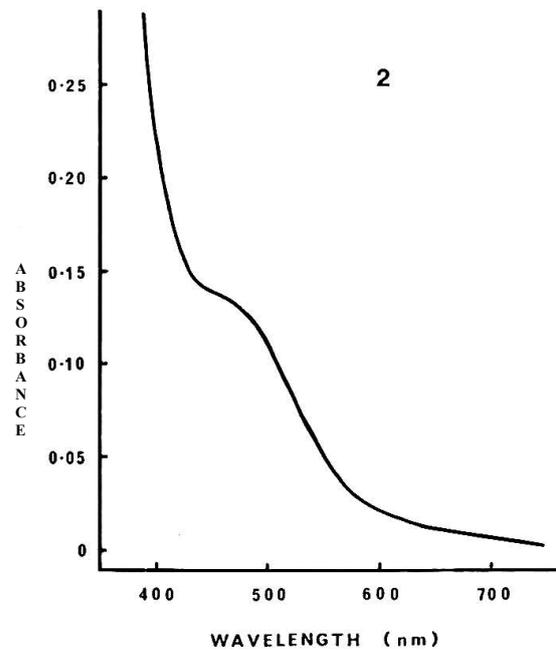


FIGURE 2
Spectrum of a wine showing pinking (from Simpson, 1977).

Simpson (1977a) found that the increase in pinking was linear up to 24 hours, reaching a peak at three days and then decreasing to 14 days. However, the reason for the shorter than three-day assay periods for pinking susceptibility used in the practice today could be that winemakers need to make a decision as quickly as possible and waiting three days for results is too long.

Simpson (1977a, 1980b) reports that, at a concentration of 75 mg/L (0.25 mL), H_2O_2 shows the most consistent results. At 75 mg/L (0.25 mL) H_2O_2 , Simpson also found that SO_2 did not influence the outcome, but at a concentration of 45 mg/L (0.15 mL) H_2O_2 , the lack of SO_2 or low concentrations of SO_2 could influence the values and therefore give a false negative to the winemaker.

Assays used in Australia, South Africa, America and Europe

The assays for pinking and various approaches used in different parts of the world are described below. Although laboratories in winemaking countries use the assay established by Simpson (1977a), there are variations in different countries adapted to best suit their final objectives.

Australia

A 100 mL clear glass screw cap bottle is labelled as 'control' and another as 'test'. The 'control' bottle is filled with wine. Forty mL of the same wine is measured into the 'test' bottle, to which 0.5 mL of 0.3% (w/v) hydrogen peroxide is added and mixed well. The 'test' sample is then placed in a dark cupboard at approximately 25°C overnight (about 12 hours). The degree of pinking of the 'test' wine is compared to that of the 'control'. In addition to visual assessment, spectral measures of the 'test' and 'control' wines can be performed at 520 nm, which gives a quantitative comparison. In this case,

the wines are filtered through a 0.45 μ m filter for assessment. A change greater than 0.050 at 500 nanometres (nm) between the control and treated sample indicates significant susceptibility to pinking (Australian Wine Research Institute [AWRI], 2020).

South Africa

According to the SASEV Methods of Analysis for Wine Laboratories (2002), a 0.072% (w/v) H_2O_2 solution (1.2 mL of 30% (w/v) H_2O_2 in 500 mL volumetric flask with distilled water) is used. A set of 5 x 25 mL sample bottles are filled with wine and additions of 0, 0.5, 0.75, 1.00 and 1.25 mL of the 0.072% (w/v) solution are done. The sample bottles are mixed gently and left for at least eight hours (the temperature and whether in a dark place or not are specified in the method). After eight hours, the samples are measured on a spectrophotometer at 500 nm, zeroed with the control sample (0 mL of H_2O_2 added) and, if the optical density (OD) is above 0.05, the wine is susceptible to pinking.

America and Europe

In both America and Europe, the method described by Simpson (1977a) is roughly followed (personnel communications). In America, 250 μ L of a 0.3% (w/v) H_2O_2 solution is added to the wine sample, while in France 125 μ L of the 30% (w/v) H_2O_2 is used. In both countries, the samples are kept in a dark cupboard for 24 hours. The specific method is not revealed by the laboratories and the personnel were not willing to part with all aspects of the methods. The spectrophotometer is zeroed with distilled water and both the control and treated samples are measured. The difference between these two is given as AU and, when the value is ≥ 0.05 , the wine is seen as having a pinking susceptibility.

Concluding remarks

Simpson (1977a) reports that if the AU is above 5, the wine shows potential for pinking susceptibility. In his research work, he multiplied the absorbance unit (AU) by 1 000 ($\times 10^3$) to get to a whole number. In all the methods, an AU of 0.05 (10 times higher than the 0.005 Simpson used in his original work) is used. It is not sure when this decision was made and for what reason, and this discrepancy has never been questioned.

The use of a 0.3% (w/v) H_2O_2 solution in Australia and America has been reported, while in South Africa it is 0.072% (w/v). In France, a 30% (w/v) undiluted solution is added to the wine sample (Table 1). Different volumes of the H_2O_2 concentrations are used by the different countries (500 μ L in 40 mL of wine sample, 250 μ L in 10 mL of wine sample, 1 000 μ L in 25 mL of wine sample and 125 μ L in 10 mL of wine sample, respectively). The final concentrations of H_2O_2 in the treatment sample differ, but could easily be worked out with the formula $C_1V_1 = C_2V_2$. This will lead to different sensitivity measurements and different absorbency units, and possibly different conclusions on whether or not a wine shows pinking susceptibility.

Another difference between the countries is that Australia uses a wavelength of 520 nm, while South Africa and France use 500 nm, as stated by Simpson (1977a). The reason for this could be that some countries scan through a wide range of spectra, i.e. 400 to 650 nm, to determine the wavelength of maximum absorbance.

Simpson (1977a) originally used a 10 mL sample bottle for his experiments. This is also the case in America and France, while in Australia a 40 mL sample bottle is used (Iland *et al.*, 2012). In South Africa, a 25 mL sample bottle is used (SASEV, 2002). The question arises if the addition of H_2O_2 from the different stock solutions will have an impact on the values for pinking susceptibility and regarding the pinking sensitivity of the wines. Would it then not be better to standardise to the original concentrations set by Simpson (1977a).

The above shows that there is no real standard for the testing of pinking susceptibility. In some cases, the waiting period is up to 24 hours, whereas in countries like South Africa it is reduced to eight hours of waiting. It is evident that there is a need to develop a standardised, shorter and more reliable method for the testing of pinking susceptibility. There is also no protocol prescribed by the International Organisation of Vine and Wine (OIV) on an assay for pinking.

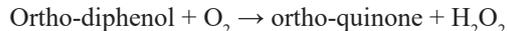
FACTORS INFLUENCING PINKING IN WHITE WINES

In a series of articles published from 1980 to 1983, Simpson and co-workers discussed different factors that could contribute to the pinking susceptibility of white wines. The factors influencing pinking in white wine are discussed on the basis of these abovementioned articles and supplemented by other authors.

The role of phenols

Singleton and Esau (1969) discussed the possibility of colourless plant phenols turning pink in an acidic medium, like wine, when the colourless anthocyanogens turn into anthocyanidins. In 1977, Simpson also stated that the spectral and chemical properties of the pink wines tested indicated that the precursors could be phenolic in origin. This started the reasoning that the oxidation of phenols could lead to pinking.

Phenolics in wine are divided into two groups. These are the flavonoids, of which the flavan-3-ols are part, and the non-flavonoids, of which the hydroxycinnamic acids and hydroxybenzoic acids are part. The flavan-3-ols consist of catechin, epicatechin, epigallocatechin and epicatechin-gallate and are found mainly in the skins and pips of grapes (Monagas *et al.*, 2005; Aron & Kennedy, 2008; Piñeiro *et al.*, 2012). The hydroxycinnamic and hydroxybenzoic acids are normally found in the fleshy parts of the grapes (Garrido & Borges, 2013; Nel, 2018). Compounds that have an ortho-diphenol grouping are highly reactive with dissolved oxygen (Garrido & Borges, 2013) to form an ortho-quinone:



These ortho-quinones are very unstable because of their highly electrophilic nature and can react in a further three ways. Firstly, the ortho-quinone can form dimers or polymers if reacting with the nucleophilic parent. Secondly, the ortho-quinone can undergo further nucleophilic additions with other nucleophiles (amino acids, glutathione, and other phenols). Thirdly, the ortho-quinone can be reduced by other reducing species, like ascorbate and other phenols, to form ortho-diphenols (Fulcrand *et al.*, 2006). All these non-enzymatic reactions are catalysed by Fe^{3+}/Fe^{2+} or Cu^{3+}/Cu^{2+} . The oxidation of these phenols leads to the browning of white wines (Fulcrand *et al.*, 2006; Garrido & Borges, 2013; Rustioni, 2017). The cause of pinking is still speculative when it comes to phenols as causative agents.

TABLE 1

The differences in the assays used in South Africa, France, America and Australia

	H_2O_2 from 30% (w/v) stock solution	Sample volume (mL)	H_2O_2 added (μ L)	Waiting period (hours)
South Africa	0.072%	25	1 000	8
Australia	0.3%	40	500	12
United States of America	0.3%	10	250	24
France	30%	10	125	24

The H_2O_2 solutions are all in % (w/v)

The attribution of several possible phenols (protocatechuic acid, catechin, epicatechin, caffeic acid, gallic acid, ethyl gallate, p-hydroxybenzoic acid, quercetin-3-rutinoside, quercetin-3-glucoside, quercetin-3-L-rhamnoside, quercetin-3-D-galactoside, cis-coutaric acid, trans-coutaric acid, m-coutaric acid, p-coutaric acid, caftaric acid, ferulic acid, fertaric acid and coumaric acid), the implication of unstable flavonoid phenols (astilbin and engeletin) and the chemical degradation of some of these procyanidins in the presence of oxygen to form anthocyanogens may lead to the appearance of a pink colour (Tobe, 1983).

The role of temperature

Simpson (1977b) states that the development of the pink colour can generally be linked to the ingress of oxygen during winemaking processes. These processes are normally critical points in which oxygen uptake plays a role, such as pump-overs, filtration, bottling, etc. The solubility of oxygen increases with a decrease in temperature (Simpson, 1980b). Oxygen solubility in wine at room temperature and atmospheric pressure is about 6.0 mL/L (8.6 mg/L) (Castellari *et al.*, 2004; Waterhouse & Laurie, 2006). This solubility increases by about 10% with a decrease in temperature (Waterhouse & Laurie, 2006). Winemakers therefore should be careful about practices in which wine temperature is kept low and the chances for oxygen uptake are high, like crushing and destemming, pressing and pumping of wine at low temperatures.

Light

Simpson (1980a) suggests that when a wine pinks in the bottle it can be exposed to direct sun or UV light for about 10 minutes to reduce the pink discoloration. UV exposure might lead to reduced pinking, but can have other negative effects. To explain the chemistry behind UV exposure, Clark *et al.* (2011) show that Fe³⁺ and light have the potential to degrade wine compounds, like tartaric acid, 3-mercaptohexanol (3MH) and 3-mercapto hexylacetate (3MHA). Different coloured glass bottles have different degrading properties and protection against UV light and the degradation of compounds. The different coloured glass bottles, with their protective abilities in increasing order, are Flint < Arctic Blue < French Green < Antique Green glass (Dias *et al.*, 2012). Light can also catalyse free radical reactions that are involved in the peroxidation step of autoxidation (Simpson, 1980b). The UV light furthermore promotes the browning of phenols in wine (Clark *et al.*, 2011; Parish-Virtue *et al.*, 2019). UV light has the ability to excite singlet oxygen, which is then able to diffuse over a large distance of 270 nm (2.7 × 10⁻⁴ mm). The singlet oxygen molecule is electrophilic, as it has a completely vacant 2pπ orbital. Therefore, the singlet oxygen molecule can react with high e-density double bonds via a six-membered ring. This results in the formation of hydroperoxide radicals (HOO•) that again assist in autoxidation (Choe & Min, 2009).

Trace metals

During non-enzymatic oxidation or chemical oxidation, H⁺ ions are transferred from a diphenol to an O₂ to form H₂O₂, but these reactions can only take place in the presence of

metal ion catalysers like Fe³⁺ and Cu²⁺. This process is mediated by the redox cycle, in which Fe³⁺/Fe²⁺ and Cu²⁺/Cu⁺ reduce oxygen to hydrogen peroxide (Oliveira *et al.*, 2011). For this reason, knowledge of the iron and copper concentration of the wine is of utmost importance, as it can have a significant impact on the autoxidation of the wine.

SO₂ concentration of the wine

During a study done by Simpson (1977a) on the effect of SO₂ on pinking susceptibility, he reduced the pH of a range of wines to pH 1. Sparging of the wine with nitrogen expelled the SO₂ from the wine. Simpson then adjusted the pH of the wines back to their normal states and added SO₂ in potassium metabisulphite form up to a free SO₂ of 60 mg/L. Two concentrations of H₂O₂, viz. 15 mL/L and 75 mL/L, were added to the wines and the pinking susceptibility was tested. With the lower concentration (15 mL/L) of H₂O₂, the pinking susceptibility was reduced proportionally with the increase in free SO₂. With the higher concentration of H₂O₂ (75 mL/L), there was a reduction in pinking at a free SO₂ of 40 mg/L. The amount of free SO₂ (concentration of about 40 mg/L) in the wine was sufficient to react with H₂O₂ to form an HSO₃⁻ anion. This will also be sufficient to prevent pinking in white wines (Simpson, 1977a).

Ascorbic acid addition as an antioxidant

Ascorbic acid is a very strong antioxidant as it reacts effectively with O₂ in the wine (Simpson, 1980a, 1980b). The H₂O₂, formed from transferring an H⁺ ion to an O₂ from ascorbic acid, is also a very strong oxidising agent (Bradshaw *et al.*, 2004, 2011; Barril *et al.*, 2016) and therefore the concentration of free SO₂ in the wine needs to be at least 40 mg/L (Simpson, 1977a). Ascorbic acid is one of the agents that works very well in preventing pinking in white wines. The addition of ascorbic acid prior to bottling may also keep the wine safe against oxidative browning in the bottle (Gibson, 2006). But there could also be a downside to the addition of ascorbic acid as it may also decrease the shelf life of the wine, with the risk of oxidative browning and even pinking (Bradshaw *et al.*, 2011; Barril *et al.*, 2016). Bradshaw *et al.* (2004) found that the molar ratio of ascorbic acid to SO₂ must be 1:1.7 to prevent oxidative browning and pinking in wines.

Wine pH

The equilibrium of molecular SO₂, bisulphite and sulphite ions in wine is pH dependent. A sulphite anion attached on the C-4 position of the anthocyanin transforms it into a colourless form. This means that, at a lower pH, more molecular SO₂ is available for the protection of the wine against oxidation (Simpson, 1980b; Abramovič *et al.*, 2015). Simpson (1977a) tested the influence of pH on pinking. Wine with a known pinking susceptibility was used to provide a pH range from 2.75 to 4.00. Pinking values were then obtained four hours after the addition of 75 mg/L H₂O₂. In a second test, samples were acidified to a pH of 1 and assayed for pinking. No significant differences were obtained in the pH range, as well as for the acidification test. This led Simpson to believe that the compound causing pinking is not a flavylum salt or its glucosides (anthocyanins). This

was confirmed by Tobe (1983), who used seven cultivars made from grapes in an experimental wine cellar in 1981, and Jones (1989), who made wine in three consecutive years (1985 to 1987). The wines that were made were treated specially for the experiments planned. Although true for monomeric anthocyanins, polymeric anthocyanins are more resistant to SO₂ bleaching and pH changes (Somers, 1971; Andrea-Silva *et al.*, 2014). During ageing and/or maturation, a polymerisation of anthocyanins takes place at the C-8 and C-6 positions, forming anthocyanin-tannin condensation reactions (Monagas *et al.*, 2005). These reactions lead to a stable polymeric anthocyanin, which therefore is resistant to decolouration by SO₂ and to pH changes (Somers, 1971).

TREATMENT OF PINKING SUSCEPTIBILITY OF WHITE WINES

Lamuela-Raventós *et al.* (2001) did a series of experiments to find the best product to remove the precursors for pinking susceptibility in white wines. Wines were divided into four lots, control wine; wine with 1 g/L PVPP, wine with 1 g/L PVPP + 0.5 g/L bentonite and wine with 1 g/L PVPP + 15 mg/L ascorbic acid. Wine treated with 1 g/L PVPP reduced pinking by 74%, the wine with 1 g/L PVPP + 0.5 g/L bentonite reduced pinking with 90%, and the wine with 1 g/L PVPP + 15 mg/L ascorbic acid reduced pinking with 98%. However, after 20 days the capacity of ascorbic acid + PVPP to reduce pinking decreased to the same levels as that of PVPP + bentonite. Tobe (1980) investigated the removal of precursors by bentonite and PVPP. He found initially observed decreases in total phenols by bentonite fining, to be ineffective after applying the Freundlich equation. PVPP was more favourable in removing the total phenols. Lamuela-Raventós *et al.* (2001) added various concentrations of ascorbic acid to a wine, i.e. 0, 15, 30, 45 and 100 mg/L. At 30 mg/L pinking was reduced, however, at 45 mg/L pinking was completely prevented. This showed that ascorbic acid is a good agent to prevent pinking susceptibility in white wine, but it could lead to oxidative browning after an extended period (Lamuela-Raventós *et al.*, 2001).

ALTERNATIVE EXPLANATIONS TO PINKING

Andrea-Silva *et al.* (2014) report the compound to cause pinking susceptibility in Siria wines to be malvidin-3-O-glucoside, which was the most abundant anthocyanin tested. Siria is a Portuguese cultivar, a widely planted Iberian variety also known as Roupeiro, Doña Blanca and Cigüente. The wine is aromatic but oxidises easily (Robinson *et al.*, 2012).

Andrea-Silva *et al.* (2014) mention that the wine used for their experiments pinked naturally. After pinking, 0.8 g of PVPP was added. The suspension was then filtered through a cheesecloth and washed with 100 mL of water and 100 mL of ethanol (95%). Thereafter, the PVPP was loaded into an empty SPE cartridge and eluted with acetonitrile and acetone, an aqueous solution of 1% HCl, ethanol and 0.1 M NH₃ in ethanol. Each fraction was kept separate. After evaporation and reconstitution with 0.2 mL of methanol and water, the samples were loaded onto an HPLC. The main compound found was malvidin-3-O-glucoside. Andrea-Silva

et al. (2014) also established that the minimum amount of total anthocyanin must be 0.3 mg/L (300 µg/L) to turn the wine a visible pink colour. This experiment was repeated in 2019 (Cosme *et al.*, 2019). Arapitsas *et al.* (2015) analysed grapes of Sauvignon blanc, Chardonnay and Riesling using a UPLC-MS/MS. They found measurable amounts of malvidin-3-O-glucoside, as well as carboxypyranomalvidin-3-O-glucoside (A-type vitisin) and pyranomalvidin-3-O-glucoside (B-type vitisin). The amounts were 55.44 µg/kg, 37.05 µg/kg and 38.99 µg/kg, respectively, for Sauvignon blanc, Chardonnay and Riesling (Arapitsas *et al.*, 2015).

In genetic analyses for anthocyanins in red and white grapes, six genes were determined in the flavonoid biosynthetic pathway. Some genes were expressed in all grapes, even where little or no anthocyanins accumulated, but an expression of the gene encoding a UDP glucose-flavonoid 3-o-glucosyl transferase (UFGT) was only detected in red grapes that synthesised anthocyanins. The analysis of the white grapes indicated that the UFGT gene was present but was not expressed (Boss *et al.*, 1996). External environmental conditions and vineyard practices therefore can switch on these genes to start the anthocyanin metabolic pathways (Boss *et al.*, 1996).

The original researcher on pinking, Dr Bob Simpson, reported that phenols (flavonoid and non-flavonoids) and not anthocyanins are the causative compound. Research on pinking in wine is thus far from over and more evidence is needed to find the colour-forming compound.

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